Omega-3 polyunsaturated fatty acids: photoprotective macronutrients

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Abstract: Ultraviolet radiation (UVR) in sunlight has deleterious effects on skin, while behavioural changes have resulted in people gaining more sun exposure. The clinical impact includes a year-on-year increase in skin cancer incidence, and topical sunscreens alone provide an inadequate measure to combat overexposure to UVR. Novel methods of photoprotection are being targeted as additional measures, with growing interest in the potential for systemic photoprotection through naturally sourced nutrients. Omega-3 polyunsaturated fatty acids (n-3 PUFA) are promising candidates, showing potential to protect the skin from UVR injury through a range of mechanisms. In this review, we discuss the biological actions of n-3 PUFA in the context of skin protection from acute and chronic UVR overexposure and describe how emerging new technologies such as nutrigenomics and lipidomics assist our understanding of the contribution of such nutrients to skin health.

Key words: lipidomics – nutrigenomics – omega-3 polyunsaturated fatty acids – photoprotection – ultraviolet radiation

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Introduction

Utilisation of bioactive nutrients for enhancement of skin health is a novel area of research. Macronutrients such as omega-3 polyunsaturated fatty acids (n-3 PUFA) are attracting attention as potential agents for maintenance of skin health and treatment of skin disorders, particularly those mediated by solar ultraviolet radiation (UVR), including sunburn, cancer, photosensitivity and photoageing (1). Skin cancers are now the commonest cancers in many white-skinned populations, and their incidence continues to rise (2). Moreover, there is increasing public awareness that most visible signs of skin ageing on exposed sites are attributable to UVR, i.e. photoageing. Eicosapentaenoic acid (EPA, 20:5n-3) is a long-chain (LC) n-3 PUFA reported to protect the skin against deleterious UVR effects, reducing UVR-induced inflammation (3–5) and indicators of photoageing and photocarcinogenesis (6–9). Omega-3 PUFA are multi-active agents that may convey photoprotection through a range of mechanisms including alterations in membrane fluidity, modification of signal transduction, transcription factor activation, modulation of oxidative stress, and production of bioactive lipid mediators. While research continues into mechanisms underlying the protective effects of LC n-3 PUFA, interest is developing into the potential effects of short-chain (SC) n-3 PUFA, such as α-linolenic (ALA, 18:3n-3) and stearidonic (STA, 18:4n-3) acids, also found in the human diet (10).

In this article, we review the state of knowledge of cutaneous n-3 PUFA biology, and the nature and mechanisms of their reported photoprotective properties. We also evaluate how the new technologies of lipidomics and nutrigenomics may further understanding of the complex relationships between UVR, cell signalling and gene expression, and their modification by n-3 PUFA.

Topical photoprotection – is it enough?

Human exposure to UVR has increased over the past 50 years, and this is largely attributable to behavioural changes relating to sun exposure, including travel to sunny holiday locations, with use of sunbeds and depletion of the stratospheric ozone contributing (11). In the future, global warming may also play a role, with warmer climates encouraging people to spend more of their leisure time outdoors (11,12). Recommended strategies for protecting skin against deleterious UVR effects include covering with clothing, avoiding midday sun, seeking shade, and topical application of sunscreens (13–16). Topical sunscreens reduce UVR effects by scattering, reflecting or absorbing radiation. However, in practice, sunscreens are not applied as thickly or evenly as in the manufacturers’ test procedures; therefore, the anticipated protection factor is not reached (17–19). This may result in a false impression of the degree of protection and then increased time spent outdoors (20). Furthermore, a significant proportion of an individual’s annual UVR exposure occurs during routine (non-holiday) activities when topical sunscreens are not typically used (21,22). While topical sunscreens can provide a high sun protection factor (SPF) against acute UVR-induced erythema, i.e. sunburn, nutrients may provide a lower level of protection against chronic, repeated UVR insults; hence, a combined approach may be optimal. Dietary nutrients acting within skin cells to modulate biological responses to UVR could provide a safe and continuous systemic approach to UVR protection, additional to the use of physical measures (9,23).

How are n-3 PUFA obtained?

Omega-3 and n-6 PUFA are regarded as essential fatty acids (EFA); human metabolism requires a dietary supply of the parent linoleic (LA, 18:2n-6) and α-linolenic (ALA;18:3n-3) acids, as well...
as some elongated and desaturated LC-PUFA including EPA and docosahexaenoic (DHA, 22:6n-3). The richest dietary sources of EPA and DHA are marine animals, with oily fish such as mackerel, sardines, herrings, and salmon containing EPA and DHA at 30–50% of tissue fatty acids (24). Negative aspects of encouraging oily fish consumption are overfishing of wild stocks and health concerns regarding heavy metal contamination and polychlorinated biphenyls (PCBs) (25,26). Potential solutions are the availability of distilled n-3 PUFA supplements, and new developments in biotechnology allowing LC n-3 PUFA enrichment of oilseed crops and micro-organisms (27). Insertion of genes encoding metabolic machinery of the n-3 pathways into the plant Arabidopsis thaliana has shown promising results, with LC n-3 PUFA reaching up to 5% of total seed fatty acids (28,29).

In contrast, SC n-3 PUFA ALA and STA are commonly found in plants and plant seed oils. The fatty acid composition of any particular food source can vary considerably and values tend to differ between reports; however, linseed, canola (rapeseed) and soyabean oils are regarded as some of the richest sources of ALA available for human consumption, containing ~53.3, 11.1 and 6.8% ALA, respectively (30). ALA is also present in walnuts and butternuts, at ~6.8% and 8.7%, and in smaller quantities (0.1–1.7%) in several other edible plants including raspberries, lettuce and peas. Sources of STA are more limited: blackcurrant seed oil contains ~3% STA, while plants of the Boraginaceae family are among the richest natural sources (31), with Echium plantagineum seed lipids containing ~13% STA (24). Echium oil-based STA-rich supplements are now marketed as vegetarian equivalents of fish oil-rich supplements, with claims of health benefits in skin (32), although published data are lacking. Genetic modification of crops has developed strains of the canola plant producing seed oils with STA levels up to 23% (33), and marine algae is another potential STA source (31).

The average daily intake of n-3 PUFA varies geographically, but in the West is typically quite low in the present day. A survey of the dietary habits of 1724 British adults over a 7-day period revealed a daily intake of 1.7–2.0 g of n-3 PUFA and 9.4–12.9 g of n-6 PUFA (34). In a study of 4884 French adults, n-3 and n-6 PUFA levels were similar to those in the UK at 1.14–1.44 g/day and 8.25–10.84 g/day, respectively (35). However, there is evidence that historically the amounts of n-3 PUFA and n-6 PUFA in the diet were balanced, i.e. with a ratio of 1:1 and with higher intakes of n-3 PUFA (36). This is proposed to be a more healthy diet than the typical Western intake that is heavily skewed towards n-6 PUFA, as in general the mediators derived from n-3 PUFA moderate the effects of the more potent, pro-inflammatory n-6 PUFA-derived mediators (36).

**Short- vs long-chain n-3 PUFA**

Parent n-3 PUFA ALA is metabolised to EPA and DHA via a series of elongation and desaturation reactions (Fig. 1). Human epidermis displays low Δ5 and Δ6 desaturase activity, and the LC forms require synthesis elsewhere, primarily in the liver, with delivery to skin through the circulation (37). However, conversion of dietary ALA to LC n-3 PUFA is relatively inefficient in mammals with only about 0.2% of plasma ALA converting to EPA (38), while the majority of dietary ALA undergoes β-oxidation for energy production (39). Several factors can influence this metabolic pathway including dietary levels of n-6 and n-3 PUFA (40).

Dietary ALA supplementation substantially increases skin phospholipid content of ALA in rodents, where it is suggested to play an active role in skin barrier function (10,41). Whether ALA accumulates in human skin in a similar manner is unclear. Stearidonic acid may act as an alternative source of EPA, bypassing the need for Δ6 desaturation of ALA. This SC n-3 PUFA acid can be directly elongated to eicosatetraenoic acid (ETA; 20:4n-3) and subsequently desaturated to EPA (Fig. 1). Although dependency on Δ5 desaturation may inhibit STA metabolism to EPA in the skin, STA could still prove more successful than ALA as a LC PUFA source. In red blood cells and plasma phospholipids, STA was approximately 2-fold more effective in increasing EPA levels than ALA (42,43). Interestingly, recent studies in hairless mice and human skin (44,45) showed that topical eicosatrienoic acid (20:3n-3) offered significant protection against UV-induced skin thickening, inflammatory cell infiltrate and transepidermal water loss (45).

While further evaluation of the application and impact of SC n-3 PUFA on the skin is required, currently the most favourable method of elevating cutaneous LC n-3 PUFA is by increasing dietary EPA content. In general, n-3 PUFA levels in human skin are very low, i.e. < 2% of total epidermal fatty acids, but this has been demonstrated to be significantly increased following dietary LC n-3 PUFA supplementation (3,7). In a human study, a 3-month course of 4 g EPA daily resulted in skin EPA levels increasing 8-fold (7). Topical application of EPA to the skin is also possible, although the relative bioavailability that can be achieved with this method is uncertain.
Cutaneous eicosanoids and other PUFA-derived mediators

Arachidonic acid (AA, 20:4n-6) and EPA are esterified in epidermal phospholipids, and released by phospholipase A2 (PLA2) isoforms, which are upregulated by UVR (46–48). Once released, AA and EPA compete for metabolism by cyclooxygenases (COX) and lipoxygenases (LOX) to produce eicosanoids, including prostaglandins (PG), thromboxanes, leukotrienes, mono- and poly-hydroxy fatty acids and lipoxins (49). These extracellular lipid-signalling molecules are involved in the regulation of various cellular processes during normal homeostasis and also play key roles during skin inflammation and tumorigenesis (49–52). Cyclooxygenase metabolism of EPA produces prostaglandins of the 3-series, e.g. PGD3, PGE3 and PGF3α, whereas LOX enzymes transform EPA to hydroxyeicosapentaenoic acids (HEPE) and 5-series leukotrienes (Fig. 2). Metabolism of AA by these enzymes produces 2-series prostaglandins (PGD2, PGE2 and PGF2α), hydroxyeicosatetraenoic acids (HETE) and 4-series leukotrienes (Fig. 2) (50,53). EPA-derived eicosanoids appear less potent than their AA-derived counterparts and tend to either act as agonists or dilute the pro-inflammatory effects of the latter (54–56). Docosahexaenoic acid can also be metabolised to a range of lipid mediators in the skin including 17-hydroxydocosahexaenoic acid (17-HoDHE), thought to be anti-inflammatory (57). Finally, a number of n-3 and n-6 cutaneous PUFA can also be metabolised by cytochrome (CYP) P450 enzymes or undergo non-enzymatic oxidation, producing an even wider variety of mediators and potential biological effects (58).

Omega-3 PUFA as regulators of transcription

Physiological effects observed as a result of changes in membrane LC-PUFA composition have been understood to be because of altered membrane fluidity and n-3/n-6 PUFA-derived eicosanoid ratios. However, LC-PUFA also modulate nuclear transcription factors such as nuclear factor-κB (NF-κB), activator protein-1 (AP-1) and sterol regulatory element-binding proteins (SREBP), and receptors such as peroxisome proliferator-activated receptors (PPAR), retinoid X receptors (RXR) and liver X receptors (LXR), composed of c-Fos and c-Jun subunits, which binds to response elements in promoter regions of genes involved in proliferation, metastasis and cellular metabolism; therefore, aberrant AP-1 activity is often observed in tumors (74). In humans, non-UVR-exposed skin constitutively expresses c-Fos with Jun-D being its

differs according to tissue, and, to date, few studies have assessed the impact of these fatty acids on UVR-induced transcriptional regulation in human skin.

**PPAR**

Both n-3 PUFA and PG are able to stimulate PPAR to bind to peroxisome proliferator response elements (PPRE) in gene promoter regions, regulating gene expression. PPAR-γ activity is upregulated by UVR in normal primary human keratinocytes (NPHK), in association with increased COX-2 expression and PGE2 synthesis, which is partially inhibited by PPAR-γ antagonists (61). UVR is proposed to increase PPAR-γ activity via generation of free radicals that oxidise glycophosphocholines (GPC) forming 1-hexadecyl 2-azaaloyl phosphatidylcholine (azPC), a potent PPAR-γ agonist (62). Using HaCat keratinocytes, Chène et al. (63) showed that both EPA and γ-linolenic acid (GLA, 18:3n-6) upregulated COX-2 expression via PPAR-γ. In contrast, PPAR-α and PPAR-β are downregulated by UVB in human skin and are associated with reduced inflammation when activated by synthetic agonists (64). Currently, it remains unclear whether some anti-inflammatory effects of LC n-3 PUFA in the skin are mediated through regulation of PPAR signalling pathways.

**NF-κB**

Nuclear factor-κB is an important transcription factor involved in epidermal homeostasis and is upregulated in response to UVR and oxidative stress (Fig. 3) (65,66). It regulates expression of many genes involved in inflammation including pro-inflammatory cytokines such as TNF-α, IL-1α, IL-1β, IL-6 and IL-8 (67). A role for NF-κB in regulating cell proliferation has also been demonstrated in transgenic mice, where repression of NF-κB increased keratinocyte proliferation and epidermal hyperplasia (68). Jin et al. (45) reported increased NFKB activity in hairless mouse skin after UVB irradiation. They also found that LC n-3-PUFA, eicosatrienoic acid, was capable of decreasing UV-induced keratinocyte proliferation and epidermal hyperplasia (68). Jin et al. (45) reported increased NFKB activity in hairless mouse skin after UVB irradiation. They also found that LC n-3-PUFA, eicosatrienoic acid, was capable of decreasing UV-induced keratinocyte proliferation and epidermal hyperplasia (68). It regulates expression of many genes involved in inflammation including pro-inflammatory cytokines such as TNF-α, IL-1α, IL-1β, IL-6 and IL-8 (67). A role for NF-κB in regulating cell proliferation has also been demonstrated in transgenic mice, where repression of NF-κB increased keratinocyte proliferation and epidermal hyperplasia (68).

Interestingly, fish oil increased the expression of antioxidant enzyme genes, manganese–super oxide dismutase (Mn–SOD) and glutathione-S-transferase (GST) by up to 4-fold in the liver, while decreasing expression of genes involved in ROS generation (70). Therefore, n-3 PUFA may inhibit NF-κB-mediated gene expression through a reduction in oxidative stress. We speculate that n-3 PUFA may modulate the expression of oxidative stress responsive enzymes in the skin, as has been reported for carotenoid antioxidants (71).

**AP-1**

Ultraviolet radiation may activate cell surface receptors directly, behaving as a ligand, or indirectly through generation of ROS, and subsequently this stimulates expression and DNA binding of transcription factor AP-1 through mitogen-activated kinase (MAPK) signalling (Fig. 3) (65,72,73). AP-1 is a heterodimer, composed of c-Fos and c-Jun subunits, which binds to response elements in promoter regions of genes involved in proliferation, metastasis and cellular metabolism; therefore, aberrant AP-1 activity is often observed in tumors (74). In humans, non-UVR-exposed skin constitutively expresses c-Fos with Jun-D being its
Omega-3 PUFA and photoprotection

The therapeutic potential of n-3 PUFA in photoprotection is summarised in Table 1.

The sunburn response

Induced by acute overexposure to UVR, this is an inflammatory reaction characterised clinically by erythema and oedema, and histologically by thickening of the stratum corneum, apoptotic epidermal cells ('sunburn cells') and dermal leucocytic infiltration (77–79). These acute responses are regulated via a network of cell-signalling pathways that are upregulated in the skin following UVR. A pathway of pivotal importance in inflammation production is the metabolism of AA by COX-2, producing pro-inflammatory mediators involved in UV inflammation. These anti-inflammatory effects have been demonstrated in human studies following supplemental fish oil (1.8 g EPA + 1.2 g DHA), where UV erythema sensitivity was significantly reduced (4,85) in association with an increase in cutaneous n-3/n-6 PUFA ratio and > 60% reduction in UV-induced skin PGE2 levels (3,4). Photoprotective effects of n-3 PUFA were confirmed in a double-blind randomised study in healthy volunteers using 4 g of purified (95%) EPA supplements; % EPA/total fatty acids in epidermal phospholipids increased 8-fold and the minimal erythemal dose (MED) increased by 36% (7). Systemic delivery of n-3 PUFA ALA has also shown some UVR protection in hairless mice, reducing erythema in association with reduced PGE2 (10). Cutaneous levels of EPA-derived eicosanoids are little explored, but a recent study found an increase in PGE2 in the sunburn response, which followed a similar time course to PGE2, with elevation at 4 h through to 72 h post-UVR (81). Omega-3 PUFA LOX products may act in an analogous manner, reducing UVR-induced n-6 PUFA-derived chemoattractants that promote an inflammatory white cell influx into the skin, although this remains to be tested (81). Reports regarding the effects of EPA on UVR-induced pro-inflammatory cytokines have been mixed, with in vitro studies in human skin cells showing a decrease in IL-8 (86) and by contrast a super-induction of IL-1β and TNF-α (87), while a human study showed no effect on IL-1β, IL-6, IL-8 and TNF-α (88).

A limited number of studies have addressed the potential of topically applied n-3 PUFA in UVR protection. Application of sardine oil (11.2% EPA, 23.6% DHA) to human skin after UVB reduced erythema by 24.5% compared to control (89). However, topical ALA proved unsuccessful in reducing UV erythema, which may be explained by the lack of epidermal desaturase activity (10).

Photosensitivity disorders

Omega-3 PUFA may also protect against certain photosensitivity disorders, i.e. conditions in which patients show abnormal reactions to UVR. In an open study of 13 patients (two men, 11 women; median age 45, range 21–81 years) with the common photosensitivity disorder polymorphic light eruption, daily supplementation with 3 g mixed n-3 PUFA (1.8 g EPA + 1.2 g DHA) for 3 months resulted in a significant decrease in sensitivity to broadband UVA (313–370 nm) papule provocation, in addition to an increase in the MED to UVB (3). In a small study of three children (all boys; ages 5–8 years) with the rare photosensitivity condition hydroa vacciniforme, supplementation with 1.5 g mixed EPA and DHA daily for 3 months resulted in all three children showing decreased erythema sensitivity to UVA, with one child additionally showing decreased erythema sensitivity to UVB. All three children also had a reduced response to lesion provocation by broadband UVA (5). The mechanisms underlying the protective effects are presently unknown, although the demonstrated reduction in UBV-induced PGE2 in the PLE study suggests that the n-3 PUFA may serve to reduce the pro-inflammatory...
and MMP-9 (gelatinase-B) after UV irradiation, also through inhibition of AP-1, which can stimulate the induction of MMP-1 (the major collagenase in humans), MMP-2 (gelatinase-A) and MMP-3 (stromelysin) (99–101). Moreover, UVB is reported to stimulate MMP-1 and MMP-3 expression via the hydroxyl radical and lipid peroxidation products (93). Addition of EPA to irradiated dermal fibroblasts in vitro inhibits expression of MMP-1⁄5 and MMP-3 (stromelysin) (99–101). In addition, topical EPA (98% EPA ethyl ester) to murine skin abrogating photoimmunosuppression have not been reported in photoprotection.

Photocarcinogenesis

This is clinically observed as deep wrinkles, reduced elasticity and uneven pigmentation (90,91), the former two changes attributed to remodelling of dermal connective tissue, i.e. the extracellular matrix (ECM), by matrix metalloproteinases (MMP). These endopeptidases are upregulated in response to UV and can degrade many components of the skin’s ECM including the major collagen and elastin networks (92–94). Both UVA and UVB are thought to increase MMP expression through generation of reactive oxygen species (95–97). However, recent work has identified that UVR may also have direct photochemical effects on components of the dermal ECM dependent on protein structure (98). Stress-associated p38 and c-Jun amino terminal kinase (JNK) are phosphorylated and activated by ROS, hence upregulating the more active form of AP-1, which can stimulate the induction of MMP-1 (the major collagenase in humans), MMP-2 (gelatinase-A) and MMP-3 (stromelysin) (99–101). Moreover, UVB is reported to stimulate MMP-1 and MMP-3 expression via the hydroxyl radical and lipid peroxidation products (93). Addition of EPA to irradiated dermal fibroblasts in vitro inhibits ERK and JNK activation, resulting in reduced c-Jun phosphorylation and decreased MMP expression (76). Similar results were observed in vivo in human skin, where topical EPA (2% w/v) inhibited expression of MMP-1 and MMP-9 (gelatinase-B) after UV irradiation, also through inhibition of p38 and JNK (6). PGE2 also induces MMP expression by macrophages and fibroblasts; modulating the balance of PG production towards the less inflammatory n-3 series may contribute to dampening of cellular responses and less damage to the surrounding ECM (102,103). In addition to the prevention of ECM degradation, topical EPA is reported to promote expression of pro-collagen I and the elastic fibre components, tropoelastin and fibrillin-1 in intrinsically aged human skin through elevated TGF-β signalling (6). Many of the effects discussed earlier may be mediated through modulation of oxidative stress by n-3 PUFA, these unstable fatty acids being speculated to act as an oxidisable buffer (4).

Photocarcinogenesis

Ultraviolet radiation is a complete carcinogen, both initiating the DNA damage that can lead to mutagenesis and promoting carcinogenesis through immunosuppression. Hairless mice fed corn oil rich in n-6 PUFA exhibited reduced latency and increased numbers of skin tumors after exposure to UVR, while in contrast, mice fed a diet supplemented with menhaden oil, rich in n-3 PUFA, exhibited increased latency and decreased tumor multiplicity (8,104,105). Associations between PUFA ingestion and skin cancer have been observed in case–control studies in humans. Hakim et al. (106) found higher intakes of n-3 PUFA were associated with reduced risk of squamous cell carcinoma, and Kune et al. (107) observed an inverse relationship between fish consumption and risk of non-melanoma skin cancer.

Black et al. (105) suggested n-3 PUFA may act during the initiation phase of tumorigenesis, as when mice were changed from an n-6- to n-3 PUFA-rich diet post-UVR, tumor numbers did not reduce. However, evidence exists that n-3 PUFA exert protective effects at the tumor promotion stage through inhibition of photoimmunosuppression (108). Ability of n-3 PUFA to protect against both local and systemic UVR-induced immune suppression has been demonstrated in mice using the contact hypersensitivity (CHS) model. Dietary EPA inhibited systemic immune suppression, resulting in a heightened CHS response to contact allergen trinitrochlorobenzene after UVB irradiation (109). In addition, topical EPA (98% EPA ethyl ester) to murine skin reduced local UVB immune suppression, evidenced by increased response to difluoronitrobenzene compared to control oil (110). As PGE2 stimulates suppressor T-cell function, these results may be attributable to relatively reduced levels of PGE2 through increased synthesis of PGE2 from EPA (111). To date, effects of n-3 PUFA in abrogating photoimmunosuppression have not been reported in humans; such studies are currently ongoing in our laboratories.

Thus, systemic n-3 PUFA may provide an approach to protect against skin cancer, potentially including those prone individuals who are immunosuppressed following organ transplantation, although this remains to be tested (112).

Safety profile

In addition to beneficial effects it is important to consider any potential adverse effects associated with long-term n-3 PUFA supplementation. Several long-term studies have been performed examining the impact of n-3 PUFA supplements in healthy people and in those with coronary heart disease, liver disease and inflammatory bowel disease, with very few adverse effects observed, besides flatulence, halitosis and loose motions (113–117). However, certain subgroups of cardiac patients may respond less favourably, as seen in a study of patients with implantable cardioverter defibrillators (ICD), where fish oil appeared to be proarrhythmic (118). It has also been suggested that co-supplementation of n-3 PUFA with antioxidants may be necessary to counteract potential increases in oxidative stress as a result of n-3 PUFA peroxidation; whereas this seems reasonable, it is not
Advancing the understanding of n-3 PUFA biology

Application of novel experimental approaches including nutrigenomics and lipidomics is anticipated to provide valuable insights into the exact roles of n-3 PUFA, including their influence on UV-irradiated skin cells. Nutrigenomics offers examination of the influence of nutrients on gene expression in health and disease. Integrated with nutrigenetic studies aiming to understand the impact of genetic variation on dietary response, and together with proteomic, metabolomic and lipidomic techniques, a more complete understanding of dietary regulation of molecular and cellular responses is becoming possible (122–126). This in turn may lead to identification of therapeutic targets in diet-related diseases and may ultimately aid development of dietary interventions based on specific nutritional requirements of individuals (127). Tools for studying genomic effects of nutrients are well established, including DNA microarrays and serial analysis of gene expression (SAGE), which allow expression of thousands of genes to be analysed simultaneously in small amounts of biological tissue/fluid. To date, few studies have utilised the microarray technique to examine the effects of PUFA on cell signalling and gene expression (128).

Lipidomics aims to provide full characterisation of lipid molecular species and their biological roles; this rapidly growing field is becoming an integral part of the multidisciplinary effort supporting systems biology, molecular mechanisms of disease pathophysiology, biomarker discovery and drug development. Lipidomics is mass spectrometry based and has resulted in a range of potent and versatile analytical methodologies for many classes of biologically important lipids, in a variety of biological matrices (129–131). The potential of this approach is exemplified in our recent reports revealing the contribution of different classes of eicosanoids to mediation of the sunburn response (81,83,132).

Conclusions

Evidence suggests that LC n-3 PUFA, particularly EPA, are capable of reducing UV-induced inflammation in human skin, in addition to potentially offering protection against photoinmunosuppression, photocarcinogenesis, photoaging and photosensitivity disorders. Ultimately, combined dietary and standard topical sunscreen measures may optimise human skin protection from sunlight. The photoprotective properties of SC n-3 PUFA are less explored but may hold potential for skin protection.

Integrated analytical approaches such as microarrays in parallel with lipidomic analyses will help elucidate the numerous pathways through which n-3 PUFA regulate cellular processes and ultimately protect human skin health.

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